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Multiomics Analysis of IMiD/CELMoD Resistant Multiple Myeloma Models Unco

Abstract #166444

Resistant Multiple Myeloma Models Uncovers Novel and Targetable Vulnerabilities in the SREBP Lipid Synthesis Pathway

Sarah A. Bird^{1,2*}, Amy Barber, PhD^{1*}, Fernando J. Sialana, PhD^{1*}, Marco P. Licciardello, PhD^{1*}, Harvey Che, PhD^{1*}, Habib Bouguenina, PhD^{1*}, Yura Grabovska, PhD^{1*}, Enze Liu, PhD^{3*}, Yakinthi Chrisochoidou, PhD^{1*}, Shannon Martin, MRes^{1*}, Jyoti Choudhary, PhD^{1*}, Brian A. Walker, PhD³, Ian Collins, PhD^{1*}, Paul Clarke, PhD^{1*} and Charlotte Pawlyn, PhD^{1,2}

¹The Institute of Cancer Research, London, United Kingdom; ²The Royal Marsden Hospital NHS Foundation Trust, London, United Kingdom; ³Melvin and Bren Simon Comprehensive Cancer Center, Division of Hematology Oncology, Indiana University, Indianapolis, IN

Background

Acquired resistance to immunomodulatory drugs (IMiDs)/cereblon (CRBN) E3 ligase modulators (CELMoDs) is a major challenge in multiple myeloma (MM) treatment. These drugs bind to the CRBN component of the CRL4^{CRBN} E3 ubiquitin ligase and designate a new set of substrates for degradation via the proteasome. Generation of resistance is frequently associated with decreased CRBN expression and this is due to genetic alteration in ~1/3rd of patients. Alternative drivers of the low CRBN state, and other mechanisms of resistance, need to be elucidated. To tackle this complex problem, MM models with acquired IMiD/CELMoD resistance were generated and multiomics analysis performed.

Methods

IMiD/CELMoD resistant human MM cell lines were generated by treating MM1s and H929 cells with lenalidomide (Len), pomalidomide (Pom) or iberdomide (Iber) at ~10x GI₅₀ concentration for ~12 weeks until resistance was achieved. Resistant cell lines and controls were characterised by whole exome sequencing (WES), RNA-Seq and proteomics. A genome-wide loss-of-function CRISPR screen (Brunello library) was carried out in Iber-resistant MM1s; gene effect scores were calculated

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with the Chronos model and compared to parental MM1s using data from the Broad Institute DepMap portal. Pathway analysis was performed using g:Profiler.

The Membrane Bound Transcription Factor Peptidase, Site 1 (MBTPS1) inhibitor PF-429242 was used to inhibit activation of the Sterol Regulatory-Element Binding Protein (SREBP) pathway and the effect on cell viability measured using CellTiter-Blue®. The Multiple Myeloma Research Foundation CoMMpass database was used to explore the correlation between mRNA expression of SREBP pathway genes in newly diagnosed patients and progression-free survival (PFS).

Results

All models were resistant to the IMiD/CELMoD with which they were generated (to 20-100x the GI₅₀ concentration) and exhibited cross-resistance to other IMiDs/CELMoDs. Functional assays showed that well-characterised effects of IMiD/CELMoD treatment, e.g. degradation of Ikaros/Aiolos, were abrogated. WES showed Pom-resistant MM1S and Pom- and Iber-resistant H929 cells had acquired mutations in *CRBN* predicted to have a high impact on function. Len-resistant H929 cells had new copy number loss at the *CRBN* locus but Len- and Iber-resistant MM1s cells had no genetic changes in *CRBN*. CRBN protein expression was reduced in all resistant lines compared to control (log2 fold changes (FCs) by proteomics ranging from -0.25 to -1.95, adj p <0.05). Together these models display diverse resistance mechanisms, reflecting the clinical picture.

Proteomic analysis of the resistant lines identified key changes in the SREBP pathway. The proteomes of the resistant lines were heterogeneous and the only pathway with common significant enrichment was SREBP/fatty acid metabolism (on analysis performed per cell line of proteins with significantly decreased expression). Stearoyl-CoA Desaturase (SCD), a key effector of the SREBP pathway, was one of only 5 proteins with significantly altered expression in all 6 cell lines compared to control. SCD had log2FCs ranging from -0.37 to -1.50 (adj p <0.05). Other key pathway members are shown in **Figure 1**.

A genome-wide CRISPR screen using Iber-resistant MM1s cells identified potential new dependencies (gene effect score <-1 in resistant cells and >-0.5 in parental cells) in 47 genes including *SCD* and *MBTPS1*. MBTPS1 is critical for activation of the SREBP pathway and demonstrated one of the largest changes in gene effect (-1.4 vs –0.3).

The activity of PF-429242, an inhibitor of MBTPS1, was explored in the resistant lines. A significant difference in GI₅₀ between Iber-resistant H929 cells and control was found (1uM and >10uM respectively) with greater activity in the resistant cells. The same pattern was observed with the other resistant H929 cell lines, but not the resistant MM1s. Other pathway inhibitors are being explored.

The expression of genes encoding SREBP pathway components was explored in the CoMMpass dataset. High *SCD* or *MBTPS1* mRNA expression was associated

with significantly worse PFS (logrank p<0.01), providing a further rationale for targeting the pathway.

Conclusions

Models representing multiple different IMiD/CELMoD resistance mechanisms have unified alterations in the SREBP pathway, highlighting a role in resistance biology and potential novel and targetable vulnerabilities.



Figure 1. Heat map showing log2FCs of SREBP pathway proteins (Reactome "Activation of gene expression by SREBF (SREBP)") in the resistant cell lines compared to control. Included proteins had a significant change (adj p <0.05) in at least 4 out of 6 lines. SCD (Stearoyl-CoA Desaturase), FASN (Fatty Acid Synthase), FDPS (Farnesyl Diphosphate Synthase), CYP51A1 (Cytochrome P450 Family 51 Subfamily A Member 1), ACACA (Acetyl-CoA Carboxylase Alpha), HMGCR (HMG-CoA Reductase).

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First Presenter

Presenter Corresponding Presenter Sarah A. Bird The Institute of Cancer Research London, United Kingdom The Royal Marsden Hospital NHS Foundation Trust London, SM2 5PT United Kingdom Email: sarah.bird@icr.ac.uk -- Will not be published

In the past 24 months, have you had any financial relationships with an ineligible company as defined above? No Signed on 08/01/2022 by *Sarah Bird*

Second Author

Amy Barber, PhD The Institute of Cancer Research London, United Kingdom **Email:** amy.barber@icr.ac.uk -- Will not be published

In the past 24 months, have you had any financial relationships with an ineligible company as defined above? No Signed on 08/01/2022 by *Amy Barber, PhD*

Third Author

Fernando J. Sialana, PhD The Institute of Cancer Research London, United Kingdom **Email:** fernando.sialana@icr.ac.uk -- Will not be published

In the past 24 months, have you had any financial relationships with an ineligible company as defined above? No Signed on 08/01/2022 by *Fernando Sialana*, *PhD*

Fourth Author

Marco P. Licciardello, PhD The Institute of Cancer Research London, SM2 5NG United Kingdom **Email:** marco.licciardello@icr.ac.uk -- Will not be published

In the past 24 months, have you had any financial relationships with an ineligible company as defined above? No Signed on 08/01/2022 by *Marco P. Licciardello, PhD*

Fifth Author

Harvey Che, PhD The Institute of Cancer Research London, SM2 5NG United Kingdom **Email:** harvey.che@icr.ac.uk -- Will not be published

In the past 24 months, have you had any financial relationships with an ineligible company as defined above? No Signed on 08/01/2022 by *Harvey Che, PhD*

Sixth Author

Habib Bouguenina, PhD The Institute of Cancer Research London, SM2 5NG United Kingdom **Email:** habib.bouguenina@icr.ac.uk -- Will not be published

In the past 24 months, have you had any financial relationships with an ineligible company as defined above? No Signed on 08/01/2022 by *Habib Bouguenina*, *PhD*

Seventh Author

Yura Grabovska, PhD The Institute of Cancer Research London, SM2 5NG United Kingdom **Email:** yura.grabovska@icr.ac.uk -- Will not be published

In the past 24 months, have you had any financial relationships with an ineligible company as defined above? No Signed on 08/01/2022 by *Yura Grabovska, PhD*

Eighth Author

Enze Liu, PhD Indiana University 980 West Walnut St. IB 544 Melvin and Bren Simon Comprehensive Cancer Center, Division of Hematology Oncology Indianapolis, IN 46202 **Email:** enzeliu@iu.edu -- Will not be published

In the past 24 months, have you had any financial relationships with an ineligible company as defined above? No

Signed on 08/01/2022 by Enze Liu, PhD

Ninth Author

Yakinthi Chrisochoidou, PhD The Institute of Cancer Research London, United Kingdom

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Email: yakinthi.chrysochoidou@icr.ac.uk -- Will not be published Alternate Email: yakinthi.chrysochoidou@icr.ac.uk -- Will not be published

In the past 24 months, have you had any financial relationships with an ineligible company as defined above? No Signed on 08/01/2022 by Yakinthi Chrisochoidou, PhD

Tenth Author

Shannon Martin, MRes The Institute of Cancer Research London, SM2 5NG United Kingdom Email: shannon.martin@icr.ac.uk -- Will not be published

In the past 24 months, have you had any financial relationships with an ineligible company as defined above? No Signed on 08/01/2022 by Shannon Martin

Eleventh Author

Jyoti Choudhary, PhD The Institute of Cancer Research London, SM2 5NG United Kingdom Email: jyoti.choudhary@icr.ac.uk -- Will not be published

In the past 24 months, have you had any financial relationships with an ineligible company as defined above? No

Signed on 08/01/2022 by Jyoti Choudhary

Twelfth Author

Brian A. Walker, PhD **Professor:** Indiana University R3 C310, 980 W Walnut St, Indiana University Division of Hematology, Melvin and Bren Simon Comprehensive Cancer Center, Division of Hematology Oncology Indianapolis, IN 46202 Fax Number: 44-208-722-4432 Email: bw75@iu.edu

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Thirteenth Author

Ian Collins, PhD The Institute of Cancer Research London, SM2 5NG United Kingdom **Email:** Ian.Collins2@icr.ac.uk -- Will not be published

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Fourteenth Author

Paul Clarke, PhD The Institute of Cancer Research London, SM2 5NG United Kingdom **Email:** paul.clarke@icr.ac.uk

In the past 24 months, have you had any financial relationships with an ineligible company as defined above? No Signed on 08/01/2022 by *Paul Clarke, PhD*

Fifteenth Author

Charlotte Pawlyn, PhD The Institute of Cancer Research London, United Kingdom The Royal Marsden Hospital NHS Foundation Trust London, SM2 5PT United Kingdom **Email:** charlotte.pawlyn@icr.ac.uk

In the past 24 months, have you had any financial relationships with an ineligible company as defined above? Yes

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